

REVIEW

Assisted reproduction technology and defects of genomic imprinting

It is estimated that approximately 1% of the newborn population of the British Isles are conceived following assisted reproduction technologies such as *in vitro* fertilisation (IVF) and intracytoplasmic sperm injection (ICSI). While the long term outcome of IVF children is mostly reassuring, some concerns remain. Specifically, recent studies have suggested a possible association between assisted conception and clinical conditions of genetic origin known as genomic imprinting defects. This has arisen from several different studies observing an excess of assisted conceptions among the rare clinical disorders of Beckwith–Wiedemann syndrome (BWS) and Angelman syndrome (AS). The numbers of such patients described in the studies to date are small but indicate a clear need for large-scale investigations to clarify the link between genomic imprinting defects and assisted conception as well as to establish the exact biological basis of any such link. In view of the strong public interest in this area of medicine, it behoves all professionals working in reproductive medicine and associated areas to be aware of these emerging data and be in a position to discuss them in as informed and responsible a manner with patients, as current data limitations permit.

Introduction

Assisted reproductive technology (ART) has enabled many infertile couples to enjoy parenthood. Since the birth of Louise Brown 27 years ago, improvements in infertility treatments have resulted in increased success rates of *in vitro* fertilisation (IVF) to a level that seems to have stabilised in recent years.¹ It is now estimated that ART accounts for greater than 1% of all births in the United Kingdom and United States, and more than 30% of all twins.² Several outcome studies have highlighted the increased complication rates in IVF-conceived children compared with the general population.^{3–5} While many of the complications are attributable to a higher number of multiple births, it has been shown that singleton IVF infants have a greater risk of low birthweight⁶ and birth defects.⁷ One of the great fears when manipulating gametes *in vitro* is the introduction of defects at the laboratory stage. Some researchers have questioned the genetic implications for offspring of intracytoplasmic sperm injection (ICSI), particularly for male infertility of genetic cause. Higher incidences of *de novo* sex chromosomal aberrations,⁸ inheritance of CF mutations and Y microdeletions^{9,10} and spermatozoal aneuploidy¹¹ have been reported following ICSI procedures. Nevertheless, most of the long term follow up data of children conceived by IVF are reassuring.¹²

Since 2002, geneticists have reported an increased incidence of IVF or ICSI conceptions among children with Beckwith–Wiedemann syndrome (BWS) and Angelman syndrome (AS). These conditions may be caused by errors of genomic imprinting. As more data emerge, a tentative link has formed between ART and imprinting defects. In the UK, Maher *et al.*¹³ found an increased frequency of IVF conceptions in BWS cohorts (4% of BWS cases were ART

conceptions, compared with 1.2% of the general population). Other researchers in the United States and France describe similarly increased association of ART among children with BWS.^{14,15} Combining the findings of these investigations results in a 4.2-fold increase in the risk of BWS for children conceived *in vitro*.¹⁶ These reports echo studies by Cox¹⁷ and Orstavik¹⁸ suggesting a link between AS and ICSI. Such alarming reports have reinforced the need for continuous surveillance of the long term outcome of children conceived with ARTs.

The possibility of statistical misinterpretation of these somewhat tentative data is heightened by different methodological approaches, as well as absence of age and control data. While this may be entirely understandable in the context of studies of very rare disorders, it leaves open the possibility of interpretational error. Nonetheless, the apparent consistent increase of actual to expected cases of genomic imprinting disorders from these several disparate sources and studies remains unexplained and confers significant potential for parental anxiety.

What is genomic imprinting?

Mendel first described single-gene inheritance in his studies of the characteristics of garden peas. The principles of Mendelian inheritance explain up to 5000 clinically significant diseases. Since Garrod and Bateson first applied Mendel's laws to inborn errors of metabolism, few exceptions to Mendelian inheritance have been observed. However, increasing knowledge of molecular detail in relation to uncommon disorders has unveiled atypical patterns of single-gene inheritance. Genomic imprinting is one example where Mendel's laws are not obeyed.

On the basis of Mendelian principles, it is assumed that genes from both parents play an equal role in development. A certain allele of an autosomal gene would be equally likely to be transmitted from a parent, of either sex, to an offspring, of either sex. However, maternal and paternal genomes are not functionally equivalent; a number of genes may have modifications, specific to the parent of origin, and are said to be imprinted. Imprinted genes show preferential expression from a specific parental allele; about 50 such genes are known and are expressed according to their sex cell lineage.¹⁹

In some genetic disorders (Table 1), the expression of the disease phenotype depends on whether the mutant allele has been inherited from the father or from the mother. Perhaps the best-studied examples in human disease are Prader–Willi syndrome (PWS) and AS. In approximately 70% of PWS cases, there is a cytogenetic deletion involving 15q11–q13 occurring on the chromosome 15 inherited from the patient's father. Thus, the genomes of these patients have genetic information on 15q11–q13 that is derived solely of maternal origin. Conversely, if there is a deletion or mutation of the maternally imprinted contribution, such that the genome contains 15q11–13 of paternal origin only, the disease will manifest as AS. Thus, the parental origin of the gene has a direct influence on the disease phenotype. This is not simply a theoretical consideration; such families exist and have been the subject of study and journal reports.²⁰

Disomy

In classical inheritance, the offspring receives an equal chromosomal contribution from each parent. Disomy refers to the unusual and rare occurrence whereby an individual receives both autosomal genes at a given locus from the same parent. Hence, that region of the chromosome is disomic—both contributions from one parent. Such a situation may have no clinical sequelae, but consider the situation of a baby disomic for a gene for which the parent was an asymptomatic carrier of an autosomal recessive mutation. Now, although only one parent is a mutation carrier, the child is affected, having homozygosity of the carrier state. This phenomenon is well reported.²¹

What if the child is disomic for a gene that is imprinted? Not surprisingly, this exact situation has been described and

paternal disomy of chromosome 15q11–13 will result in no maternal contribution and a presentation of AS. Conversely, maternal disomy at this same region causes PWS; as in the deletional form, there is no paternal contribution.

How are genes imprinted?

At an imprinted locus, only one allele is active and the inactive one is marked epigenetically, that is, there is a stable alteration in DNA other than the sequence itself. Epigenetic modifications include histone acetylation, cytosine methylation or both and essentially alter chromatin organisation. Methylation is one of the best-studied epigenetic modifications of DNA and all imprinted genes show differences in methylation patterns between maternal and paternal alleles. Loss of imprinting can involve hypomethylation or hypermethylation, depending on the gene.²²

Imprinting occurs at two stages; gametogenesis and embryonic development. Imprints are established during the development of the germ cells. The imprinted genes initially undergo demethylation as primordial germ cells migrate along the genital ridge to the fetal gonad. Subsequently during gamete maturation, methylation is reestablished by DNA methyltransferases that specifically target one of the two parental alleles for silencing. Further changes occur after fertilisation. Firstly there is genome-wide demethylation, which is then followed by passive remethylation in the zygote genome. However, methylation marks on imprinted genes are protected from demethylation so that parental imprints are preserved in the developing embryo. Normal embryogenesis cannot proceed without this dynamic reprogramming of the epigenome at different stages.

Imprinted genes in development

Genomic imprinting represents a form of gene regulation. Many imprinted genes are known to play important roles in fetal growth and development, and also in tumour suppression. Moll *et al.*²³ reported an increase in retinoblastoma in IVF children and this is reminiscent of other reports of cancer in children born after assisted reproductive technologies.^{24,25} Animal studies have shown that loss of imprinting at the maternal *Igf2* gene is associated with overgrowth and large offspring syndrome,²⁶ a condition some researchers have compared with BWS in humans where organ overgrowth also occurs.

Beckwith–Wiedemann and Angelman syndromes

BWS is a rare disorder with an incidence of 1 in 14,500 live births.²⁷ The cardinal features are fetal macrosomia, abdominal wall defects, neonatal hypoglycaemia and

Table 1. Conditions associated with genomic imprinting defects.

PWS
AS
BWS
Russell–Silver syndrome
Wilm's tumour
Osteosarcoma
Bilateral retinoblastoma
Rhabdomyosarcoma

macroglossia (Fig. 1). In utero there may be a large and thickened placenta, polyhydramnios, a long umbilical cord and a fetus measuring large for gestational age. Additional, more variable features include hemihypertrophy, ear pits and creases, renal anomalies and facial nevus flammeus. BWS children are at increased risk of developing embryonal tumours, especially Wilm's tumour for which condition screening is recommended in this group.

BWS can occur by a variety of mechanisms (Table 2), but in the majority of cases imprinted genes from chromosome 11p15.5 have been implicated. About 60% of sporadic cases have epigenetic changes of methylation causing alterations in the expression of the paternally expressed alleles IGF2 and KCNQ1OT, or maternally expressed genes such as H19 and CDKN1C.²⁸ Twenty percent of cases are attributable to uniparental disomy where the inheritance of two alleles from the same parent causes loss of imprinting of a normally imprinted gene. These data from sporadic cases of BWS contrast with the results of molecular analysis of the ART-associated cases; of 19 cases found, 14 cases were tested; all were positive for hypomethylation of KCNQ1OT, and all were negative for UPD^{13–15} (Table 3).

AS is a neurogenetic disorder characterised by severe mental retardation, delayed motor development, poor balance, jerky movements, absence of speech and happy disposition (Fig. 2). Sporadic cases of AS are linked with a loss of function of the maternal allele of *UBE3A* on chromosome 15 resulting from a deletion (70%), a point mutation, uniparental disomy (2–3%) or an imprinting defect (7–9%) (Table 4). This is in contrast to findings by Cox *et al.*¹⁷ and Orstavik *et al.*¹⁸ where, in all three cases conceived by ICSI, AS was due imprinting defects caused by aberrant methylation (Table 3); a mechanism which, in the whole of the AS population, is thought to account for only 5% of cases.²⁹

More recently, Horsthemke *et al.*³⁰ reported their study among 79 patients in the German AS Support Group.³¹



Fig. 1. The macroglossia of BWS, a very classical characteristic clinical sign, is demonstrated.

Table 2. Molecular causes of BWS (adapted from Maher²²).

40%	Imprinting centre 2 defect (epimutation, loss of maternal methylation)
20%	Uniparental disomy—paternal, of chromosome 11p15.5
5%	Imprinting centre 1 defect (gain of maternal methylation)
5–10%	Mutation of CDKN1C gene on 11p15 (accounts for 40% of familial cases of BWS) ²⁸
2%	Paternal duplication or maternal rearrangement of chromosome 11p15.5
1%	Imprinting centre (IC1 or IC2) deletion
22–27%	Mechanism undetermined

Sixteen were born to subfertile couples and 4 of these 16 were due to a sporadic imprinting defect. The relative risk of AS was significantly increased among patients conceived by ICSI and among those treated by hormonal measures. This observation extends the possible risk of imprinting defects to other modalities of treatment for infertility beyond ICSI. The authors concluded that genetic predisposition, combined with superovulation, rather than ICSI, increases the risk of conceiving a child with an imprinting defect.

From these sets of observations in patients with BWS and AS, it is clear that IVF conceptions are statistically over-represented. That the molecular-specific subgroups of AS and also of BWS represent so uncommon a molecular mechanism adds up to serious concerns on the possible effects of assisted reproductive techniques on epigenetic mechanisms and imprinting. Such concerns are compounded by animal observations that overgrowth in the IVF-conceived offspring correlates with loss of methylation in the maternally imprinted IGF2 receptor gene.²⁶ In a similar vein, studies of pre-implantation mouse embryos have shown that embryo culture conditions such as presence or absence of calf serum can influence the expression and methylation status of imprinted genes.³²

Mechanism of imprinting defect in ART

It is unknown at which step, or steps, the association of ART with imprinting defects occurs. Because imprinted genes are functionally haploid, they may be vulnerable to mutations or epimutations when placed in an abnormal *in vitro* environment. Curiously, so far only maternal loss of imprinting has been described in the ART-associated cases although data is limited to small numbers.

Possibilities for the introduction of a genetic error include the following:

1. Elimination of natural selection. ICSI bypasses natural selection and may overcome intrinsic barriers to the fertilisation of abnormal gametes (e.g. those with defective imprinting) of either sex. High frequencies of cytogenetic abnormalities are detected in oocytes generated from IVF.^{33,34} It should also be acknowledged that epigenetic

Table 3. Summary of studies of imprinting disorders after ART.

Syndrome	Reference	Molecular defect	No. of ART cases	No. analysed molecularly	ART
BWS	DeBaun <i>et al.</i> ¹⁴	4/6 KCNQ1OT hypomethylation 1/6 KCNQ1OT hypomethylation, and H19 hypermethylation	7	6	IVF and ICSI
	Maher <i>et al.</i> ¹³	2/2 KCNQ1OT hypomethylation	6	2	IVF and ICSI
	Gicquel <i>et al.</i> ¹⁵	6/6 KCNQ1OT hypomethylation	6	6	IVF and ICSI
AS	Cox <i>et al.</i> ¹⁷	2/2 Sporadic imprinting defect at 1C	2	2	ICSI
	Orstavik <i>et al.</i> ¹⁸	Sporadic imprinting defect at 1C	1	1	ICSI
RB	Moll <i>et al.</i> ²³	1/5 coding mutation at RB1	5	5	IVF and ICSI

errors could be a significant cause of the underlying infertility rather than a consequence of the treatment.

- Abnormal oocyte activation. ICSI introduces the sperm acrosome and digestive enzymes into the ooplasm and may disturb intracellular homeostatic mechanisms.
- Removal of cumulus cells. Oocytes are denuded of their surrounding cumulus cells prior to micromanipulation. The cumulus complex may have a role in maintaining the oocyte in meiotic arrest, and its removal may be associated with the oocyte's irreversible commitment to undergo germinal vesicle breakdown.³⁵ By altering the meiotic competence of the oocyte, such manipulation may leave open the possibility of defective genomic imprinting.
- Mechanical disruption of the oocyte. The injecting needle may cause mechanical disruption of intracellular structures such as the meiotic spindle.

Imprinting defects have been described following IVF with and without ICSI suggesting that the error occurs in a step or steps common to both procedures. Indeed, to date there is no convincing data that ICSI entails more dangers than IVF. Common and potentially causative steps include the following:



Fig. 2. A typical case of AS is shown. Note the absence of dysmorphic features in this developmentally delayed, ataxic, speechless boy.

- Culture conditions of the ovum. Factors such as the length of exposure to specific media and growth factors therein may alter oocyte maturation. Studies of mouse embryos have demonstrated that H19 methylation and expression can be altered by the culture medium used.³² Niemitz and Feinberg³⁶ hypothesise that the methionone content of media may be a critical factor influencing methylation changes.

It seems that *in vitro* effects are greater on the oocyte, consistent with the relatively late completion of imprinting in oogenesis. Testicular sperm extraction (TESE) procedures collect immature spermatozoa, but the paternal imprinting process seems to be complete at this stage. This would point to potential extra risks with *in vitro* maturation (IVM) of oocytes, but so far there have been no reports of AS or BWS in the approximately 300 infants born to date.

- Embryo culture. Data from animal studies have demonstrated aberrant maternal allele methylation occurring in embryos cultured *in vitro*,³² so it remains possible that prolonged embryonal cell culture *per se* might predispose to abnormal methylation. This may have implications for the practice of extending culture to blastocyst stage. *In vitro* blastocyst culture, being an abnormal environment for this stage, may potentially represent an embryonic imprinting insult.
- Subfertility and superovulation. It is possible that subfertility itself and imprinting defects share a common aetiology, and that superovulation rather than *in vitro* gamete manipulation may increase the risk of conceiving a child with defect of genomic imprinting.³⁰

Table 4. Molecular causes of AS (adapted from Clayton-Smith and Laan²⁹).

70%	<i>De novo</i> maternal deletion
5–10%	<i>UBE3A</i> mutation
2%	Uniparental disomy (paternal)
2%	Imprinting centre deletion
2%	Imprinting centre defect without deletion
10–15%	None identified

The current position

There are strong circumstantial observations that suggest a cause-and-effect relationship between assisted conception and clinical conditions caused by imprinting mutations. Such data as currently exist arise from patient cohorts with two specific conditions caused by imprinting defects and show an unexpected over-representation of assisted reproduction conceptions among these patient groups. The concerns that ART may predispose to imprinting defects in offspring gain further weight with consideration of animal studies germane to this area.

Infertility specialists need to be aware of these developing issues to be well enough informed to address the legitimate concerns of their patients. It must be recognised that many patients are looking outside the profession for information nowadays and specialists need to have in mind the data available from such resources as the Human Embryology Authority Web site (www.hfea.gov.uk) and the American Academy of Paediatrics (www.aap.org) when addressing concerns raised by patients. While the patient's autonomy in decision-making is respected, an appreciation of all risks is necessary to ensure that infertile couples receive the information sufficient to make informed choices. Physicians also bear a responsibility to the health of future children and potential health risks must always be considered. The ultimate aim of infertility treatment is not just a positive pregnancy test, rather the birth of a healthy baby to healthy parents.

The absolute numbers of imprinting defects associated with IVF and ICSI are small and unlikely to deter any would-be patients from undergoing the treatment. Nevertheless, the emerging data are of concern and highlight the need for further investigation. Foremost among immediate studies should be a widespread review of ART prevalence among AS and BWS cases tested and identified in approved laboratories nationally and internationally. Concurrent with this, studies are needed to look at ART populations of children and identify if there is evidence for increased prevalence of AS and BWS in such children. Of necessity these studies will need to be large-scale, multicentred and take account of children with 'other' defects not known currently to be due to imprinting problems.

Infertility specialists, obstetricians, paediatricians and geneticists commonly work in separate environments, so that in the past issues pertaining to the short and long term effects of assisted conception have been reviewed in isolation. Increased cross-specialty co-operation, which would be greatly enhanced by appropriate research funding, and the promotion of follow up studies of patients born of assisted conception are clearly the means to definitively address the current impasse. The issues raised here are not to be dismissed as 'academic'. They are real and absolute and will impact on clinicians working in infertility and on their patients.

Acknowledgements

The authors wish to acknowledge the support of their work by the Children's Medical and Research Foundation at Our Lady's Hospital for Sick Children, Dublin.

Cathy Allen,^a William Reardon^b

^a*Human Assisted Reproduction Ireland, Rotunda Hospital, Dublin, Ireland*

^b*National Centre for Medical Genetics, Our Lady's Hospital for Sick Children, Crumlin, Dublin, Ireland*

References

1. Assisted reproductive technology in the United States: 1996 results generated from the American Society for Reproductive Medicine/Society for Assisted Reproductive Technology Registry. *Fertil Steril* 1999;**71**:798–807.
2. Assisted reproductive technology in Europe, 2001. Results generated from European registers by ESHRE. *Hum Reprod* 2005;**20**(5):1158–1176.
3. FIVNAT (French In Vitro National). Pregnancies and births resulting from in vitro fertilization: French national registry, analysis of data 1986 to 1990. *Fertil Steril* 1995;**64**(4):746–756.
4. Dhont M, De Sutter P, Ruysinck G, Martens G, Bekaert A. Perinatal outcomes of pregnancies after assisted reproduction: case-control study. *Am J Obstet Gynecol* 1999;**181**(3):688–695.
5. Births in Great Britain resulting from assisted conception, 1978 to 1987. MRC Working Party on Children Conceived by In Vitro Fertilisation. *BMJ* 1990;**300**:1229–1233.
6. Schieve LA, Meikle SF, Ferre C, Peterson H, Jeng G, Wilcox L. Low and very low birth weight in infants conceived with the use of assisted reproductive technology. *N Engl J Med* 2002;**346**:731–737.
7. Hansen M, Kurinczuk JJ, Bower C, Webb S. The risk of major birth defects after intracytoplasmic sperm injection and in vitro fertilization. *N Engl J Med* 2002;**346**:725–730.
8. Bonduelle M, Camus M, DeVos A, et al. Seven years of intracytoplasmic sperm injection and follow-up of 1987 subsequent children. *Hum Reprod* 1999;**14**(Suppl 1):243–264.
9. Van der Ven K, Messer L, van der Ven H, et al. Cystic fibrosis mutation screening in healthy men with reduced sperm quality. *Hum Reprod* 1996;**11**:513–517.
10. Pryor JL, Kent-First M, Muallem A, et al. Microdeletions in the Y chromosome of infertile men. *N Engl J Med* 1997;**336**:534–539.
11. Martin RH. The risk of chromosomal abnormalities following ICSI. *Hum Reprod* 1996;**11**:924–925.
12. Sutcliffe A, Taylor B, Saunders K, Thornton S, Lieberman B, Grudzinskas J. Outcome in the second year of life after in-vitro fertilization by intracytoplasmic sperm injection: a UK case-control study. *Lancet* 2001;**357**:2080–2084.
13. Maher ER, Brueton LA, Bowdin SC, et al. Beckwith-Wiedemann syndrome and assisted reproductive technology (ART). *J Med Genet* 2003;**40**:62–64.
14. DeBaun M, Niemitz E, Feinberg A. Association of in vitro fertilisation with Beckwith-Wiedemann syndrome and epigenetic alterations of LIT1 and H19. *Am J Hum Genet* 2003;**72**:156–160.
15. Gicquel C, Gaston V, Mandelbaum J, Siffroi J, Flahault A, Le Bouc Y. In vitro fertilisation may increase the risk of Beckwith-Wiedemann syndrome related to the abnormal imprinting of the KCNQ1OT gene. *Am J Hum Genet* 2003;**72**:1338–1341.
16. Gosden R, Trasler J, Lucifero D, Faddy M. Rare congenital disorders, imprinted genes, and assisted reproductive technology. *Lancet* 2003;**36**(9373):1975–1977.

17. Cox GF, Burger J, Lip V, et al. Intracytoplasmic sperm injection may increase the risk of imprinting defects. *Am J Hum Genet* 2002;**71**: 162–164.
18. Orstavik KH, Eiklik K, Van Der Hagen CB, et al. Another case of imprinting defects in a girl with Angelman syndrome who was conceived by intracytoplasmic sperm injection. *Am J Hum Genet* 2003;**72**:218–219.
19. Morison I, Reeve A. A catalogue of imprinted genes and parent-of-origin effects in humans and animals. *Hum Mol Genet* 1998;**7**:1599–1609.
20. Hulten M, Armstrong S, Challinor P, et al. Genomic imprinting in an Angelman and Prader–Willi translocation family. *Lancet* 1991;**2**: 638–639.
21. Gelb BD, Willner JP, Dunn TM, et al. Paternal uniparental disomy for chromosome 1 revealed by molecular analysis of a patient with pycnodysostosis. *Am J Hum Genet* 1998;**62**:848–854.
22. Maher E. Imprinting and assisted reproductive technology. *Hum Mol Genet* 2005;**14**(Suppl 1):R133–R138.
23. Moll AC, Imhof SM, Cruysberg JRM, et al. Incidence of retinoblastoma in children born after in vitro fertilization. *Lancet* 2003;**361**:309–310.
24. Kramer S, Ward E, Meadows AT, et al. Medical and drug risk factors associated with neuroblastoma: a case control study. *J Natl Cancer Inst* 1987;**78**:797–804.
25. Steensel-Moll HA, Walkenburg HA, Vanderbroucke JP, et al. Are maternal fertility problems related to childhood leukaemia? *Int J Epidemiol* 1985;**14**:555–559.
26. Young LE, Fernandes K, McEvoy TG, et al. Epigenetic change in IGF2R is associated with fetal overgrowth after sheep embryo culture. *Nat Genet* 2001;**27**:153–154.
27. Sutcliffe A. Intracytoplasmic sperm injection and other aspects of new reproductive technologies. *Arch Dis Child* 2000;**83**:98–101.
28. Diaz-Meyer N, Yang Y, Sait SN, Maher ER, Higgins MJ. Alternative mechanisms associated with silencing of *CDKN1C* in Beckwith–Weidemann syndrome. *J Med Genet* 2005;**42**:648–655.
29. Clayton-Smith J, Laan L. Angelman syndrome: a review of the clinical and genetic aspects. *J Med Genet* 2003;**40**:87–95.
30. Horsthemke B, Gross S, Katalivic A, Sutcliffe A, Varon R, Ludwig M. Subfertility is associated with an increased risk of conceiving a child with an imprinting defect. *Am J Hum Genet* 2004 [Abs 113. P.40 only].
31. Ludwig M, Katalinic A, Groß S, Sutcliffe A, Varon R, Horsthemke B. Increased prevalence of imprinting defects in patients with Angelman syndrome born to subfertile couples. *J Med Genet* 2005;**42**:289–291.
32. Khosla S, Dean W, Brown D, Reik W, Feil R. Culture of preimplantation mouse embryos affects fetal development and the expression of imprinted genes. *Biol Reprod* 2001;**64**:918–926.
33. Edirisinghe W, Murch A, Junk S, Yovich J. Cytogenetic abnormalities of unfertilized oocytes generated from in vitro fertilization and intracytoplasmic sperm injection: a double-blind study. *Hum Reprod* 1997;**12**:2784–2791.
34. Voullaire L, Wilton L, McBain J, Callaghan T, Williamson R. Chromosome abnormalities identified by comparative genomic hybridization in embryos from women with repeated implantation failure. *Mol Hum Reprod* 2002;**8**:1035–1041.
35. Racowsky C, Satterlie RA. Metabolic, fluorescent and electrical coupling between hamster oocytes and cumulus cells during meiotic maturation in vivo and in vitro. *Dev Biol* 1985;**108**(1):191–202.
36. Niemitz E, Feinberg A. Epigenetics and assisted reproduction: a call for investigation. *Am J Hum Genet* 2004;**74**:599–609.